## FIRST SELECTIVE LINBAB BUTMIENE-PUNCTIONALIZBD DIENE CODIMERIZATION CATALYZED BY NICKEL COMPLEXES

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*Summary* : *Functionalized dienes are codimerized with hutadiene in the presence of Ni'-AMP\* catalysts to produce selectively linear trienic esters. In the case oj methyl sorbate, some asymmetric induction is observed in the product.* 

Butadiene-functionalized diene cyclodimerization over nickel catalysts is a well established process, by which the use of phosphines as ligands provides the synthesis of cyclooctadiene derivatives<sup>1-3</sup>. In each case however, the reaction rates were low. We have developed the concept of ancillary bifunctional aminophosphinite (AMP) ligand where the secondary amino group can interact with a nickel complex intermediate to produce selectively  $1,3,6$ -octatrienes 1 under very mild conditions from butadiene  $^{4}$ . We wish now to report the results obtained during codimerization of methyl-2,4 pentadienoate 2 and methyl sorbate 3 with butadiene on this catalytic system. The codimerization was carried out with bis(cyclooctadiene) nickel as the source of Ni" and D-EPHOS NH, a ligand obtained by reaction between (lS,2R) ephedrine and dimethylaminodiphenylphosphine  $(eq.1)^4$ .

 $MeHNCH(Me)CH(Ph)OH + PPh<sub>2</sub>NMe,$  We  $MeHN(Me)CH(Ph)OPPh$ , (eq.1)

In a typical experiment, Ni(COD)<sub>2</sub>(110mg;0.4mg) is introduced in a Schlenk tube equiped with a Teflon tap, The ligand (140 mg;0,4 mmol) is added together with the ester in a toluene solution (7ml),followed by introduction of butadiene.The conversion are calculated by GC. The compounds structures were determined by proton and  $^{13}$ C NMR spectroscopies as well as by IR  $^5$ . These analyses are consistent with following general reaction :



The results summarized in the table are interesting in several respects:they show first that linear trienic esters can be easily prepared by this way and hence provide



useful starting materials for synthetic application. Secondly, they show how the use of a bifunctional ligand can enhance the chemoselectivity into linear products, even in cases where this process was unknown (methyl sorbate dimerization). It has to be noticed at last that codimers **5a** and **5b** have a prochiral carbon atom : the use of the chiral D-EPHOSNH as ligand has given some asymmetric induction in 5a  $(\lceil \alpha \rceil_{\Pi}^{20} = +1.$  $6(C=1.0,~CHCl<sub>3</sub>)$  and **5b**  $((a)<sub>n</sub>$ <sup>20</sup> = +30  $(C=1.0, ~CHCl<sub>3</sub>)$ .

Further experiments are under way in order to analyse the enantioselectivity observed in this chemoselective reaction.

## REFERENCES AND **NOTES**

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 $5 - 4a$   $^{1}$ H NMR (60MHZ, CDCl<sub>3</sub>) : 2.3-3.15(m, 4H) ; 3.7(S.3H) ; 5.3-7(m, 7H).

IR: 2980, 2960, 2830, 1740, 1600, 1430, 1270, 1035, 980, 895cm<sup>-1 13</sup>C NMR; 172.2  $(C=0)$ , 148.3 (C3), 135,4 (C4), 130 (C6), 132.9 (C7), 137.9 (C8), 117.7 (C9), 37.8 (C2), 31.7 (C5), 51.3 (OCH<sub>3</sub>). <u>4c</u> <sup>1</sup>H NMR (60 MHz, CDC1<sub>3</sub>) : 2.11(m, 2H,  $J_{6-5} = J_{6-7} = 5.1$ ) ; 3.  $7(S.3H)$ ; 1.65  $(d, 3H, Jg-g = 3.18)$ , 5.1 - 6.51(m, 6H). IR : same as  $4a$  except C=0 = 1680  $\underline{vs}$  1740 cm<sup>-1</sup> ;  $^{13}$ C NMR : 168.0 (C=0), 122.5 (C<sub>2</sub>), 124 (C<sub>3</sub>), 129.2 (C<sub>4</sub>), 130.8  $(C_5)$ ,128.6  $(C_7)$ , 122.9  $(C_8)$ , 35.2  $(C_6)$ , 17.73  $(C_9)$ , 51.6  $(OCH_3)$ . 5a - <sup>1</sup>H NMR (60 MHZ, CDC1<sub>3</sub>) **:** 1.1(d,  $J_{5-10} = 6.53$  ; 3H) ; 2.15-2.3(m, 3H) ; 3.68 (S, 3H) ; 5.1-6.7(m, 7H). IR : 2980, 2960, 2830, 2820, 1734, 1600, 1430, 1310, 1270, 1175, 1000.  $^{13}$ C NMR : 172.29  $(C=0)$ ; 138.42  $(C_3)$ ; 138  $(C_4)$ ; 137.09  $(C_6)$ ; 120.57  $(C_7)$ ; 129.64  $(C_8$ ; 115.46  $(C_9)$ ; 51.67 (OCH<sub>3</sub>) ; 37.8 (C<sub>2</sub>) ; 39.13 (C<sub>5</sub> ; 19.87 (C<sub>10</sub>). [a]<sup>20</sup><sub>D</sub> = +1.6(C 1.0 ; CHC1<sub>3</sub>). <u>5b -</u>  $\frac{1_H \text{ NMR}}{4}$  (60 MHZ, CDC1<sub>3</sub>) : 1.07 (d, J<sub>5-10</sub> = 6.53 ; 3H) ; 2.95-3.05 (m, 3H) ; 3.7(S, 3H) ; 5-6.5 (m, 7H).<u>IR</u> same as  $5a$ .  $^{13}$ C NMR : 172.11 (C=0) ; 138.0 (C3) ; 137.2 (C<sub>4</sub>) ; 137 (C<sub>6</sub>) **;120.1** (C<sub>7</sub>) ; 129 (C<sub>8</sub>) ; 117.58 (C<sub>9</sub>) ; 51.33 (OCH<sub>3</sub>) ; 35.13 (C<sub>12</sub>) ; 37.52 (C<sub>5</sub>) ; 20.7  $(C_{10})$  [a]<sup>20</sup> = +30 (C 1.0 ;CHCl<sub>3</sub>). **<u>5c</u>** <sup>1</sup>H NMR (60MHZ, CDCl<sub>3</sub>) : 1.59 (5.3H) ; 1.86 (d,  $J_{8-9}$  = 5.15 ; 3H) ; 3.15 (d, $J_{6-7}$  = 5.86 ; 2H) ; 3.68 (S.3H) ; 5-6.37 (m,5H). IR : Same as <u>5a</u> and <u>5b</u> except  $C=0 = 1700$  <u>vs</u> 1734 cm<sup>-1</sup>. <sup>13</sup>C NMR : 167.91 (C=0) ; 121.73 (C<sub>2</sub>) ; 122.51 (C<sub>3</sub>) ;136.71 (C<sub>4</sub>) ; 140.11 (C<sub>5</sub>) ; 129.10 (C7) ; 122.01 (C<sub>8</sub>) ; 51.73 (OCH<sub>3</sub>) ; 17. 15  $(C_q)$  ; 36.51  $(C_6)$  ; 18.15  $(C10)$ .

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